

THE RIGHT FORMULATION – AND HOW TO GET THERE

The objective of drug formulation is to develop a product with the correct amount of drug in the right form, and to maintain its chemical and biological integrity for delivery at or over the proper time, at the proper rate, and in the desired location. SRI International's approach to drug formulation is customised according to the client's needs and the stage of development. This article by Gita Shankar, PhD, Director of Formulations R&D, Pharmaceutical Sciences at SRI, describes the strategy that the company uses to develop an appropriate formulation.

Formulators start their task by gaining an understanding of the bulk drug's physical and chemical properties. Once the pharmaceutical profile of the drug has been determined, the appropriate route of administration can be identified and designed. Next we develop preclinical formulations, which overcome the drug's innate deficiencies. For example, a poorly soluble drug might require solubilising additives, and a poorly bioavailable drug might require a permeability enhancer.

Additional excipients are selected to overcome potential problems in processing, manufacturing and stability. Bench studies are performed on pilot batches to establish the efficacy, manufacturability and stability of dosage forms. The final formulation design is optimised to take into account the pharmacokinetic properties of absorption, distribution, metabolism, and excretion.

These steps in the drug development process are common to all routes of administration. However, this article covers only the development of oral dosage formulations.

Oral formulation has been the preferred and most common route of delivery around the globe owing to its ease of administration and good patient compliance. And from a drug development and manufacturing perspective, an oral formulation offers superior stability compared with intravenous formulations. Developing an oral formulation is by no means an easy task because each drug substance is a different entity with different characteristics.

To standardise oral formulation development, the US FDA published the biopharmaceutical classification system (BCS) guidance in 2000, which formed the basis of the scientific framework used for classifying drug

substances based on their aqueous solubility and intestinal permeability.¹

The biopharmaceutical classification system was developed primarily in the context of immediate release (IR) solid oral dosage forms. It is a drug development tool that allows estimation of the contributions of three major factors – dissolution, solubility and intestinal permeability – that affect oral drug absorption from IR solid oral dosage forms. The classification is associated with a drug dissolution and absorption model that identifies the key parameters controlling drug absorption as a set of dimensionless numbers: the absorption number, the dissolution number and the dose number.

According to the BCS, there are four classes of drug substances based solely on their solubility and intestinal permeability:

Class I: High Solubility – High Permeability Class II: Low Solubility – High Permeability Class III: High Solubility – Low Permeability Class IV: Low Solubility – Low Permeability

Class I drugs exhibit a high absorption number and a high dissolution number. The rate-limiting step is drug dissolution; if dissolution is very rapid, then the gastric emptying rate becomes the rate-determining step. Metoprolol, diltiazem, verapamil, and propranolol are examples of Class I drugs.

Class II drugs have a high absorption number but a low dissolution number. *In vivo* drug dissolution is then a rate-limiting step for absorption except at a very high dose number. The absorption rate for Class II drugs is usually slower than the rate for Class I drugs and absorption occurs over a longer period. Phenytoin, danazol,



Figure 1: High Throughput Liquid Handling System.



Gita Shankar, PhD
Director of Formulations R&D,
Pharmaceutical Sciences
T: +1 650 859 2000
F: +1 650 859 3041
E: gita.shankar@sri.com

SRI International 333 Ravenswood Avenue Menlo Park CA 94025 United States

www.sri.com/biosciences

Binary Combinations (1:1)		Ternary Combinations (1:1:1)	
Solvent Combination	Abbreviation	Solvent Combination	Abbreviation
Ethanol / Water	Eth Wat	Ethanol / Water / PEG400	Eth Wat PEG
Ethanol / PEG400	Eth PEG	Ethanol / Water / Propylene Glycol	Eth Wat PG
Ethanol / Propylene Glycol	Eth PG	Ethanol / Water / Tween80	Eth Wat Twe
Ethanol / Tween80	Eth Twe	Ethanol / Water / Glycerin	Eth Wat Gly
Ethanol / Glycerin	Eth Gly	Ethanol / PEG400 / Propylene Glycol	Eth PEG PG
Water / PEG400	Wat PEG	Ethanol / PEG400 / Tween80	Eth PEG Twe
Water / Propylene Glycol	Wat PG	Ethanol / PEG400 / Glycerin	Eth PEG Gly
Water / Tween80	Wat Twe	Ethanol / Propylene Glycol / Tween80	Eth PG Twe
Water / Glycerin	Wat Gly	Ethanol / Propylene Glycol / Glycerin	Eth PG Gly
PEG400 / Propylene Glycol	PEG PG	Ethanol / Tween80 / Glycerin	Eth Twe Gly
PEG400 / Tween80	PEG Twe	Water / PEG400 / Propylene Glycol	Wat PEG PG
PEG400 / Glycerin	PEG Gly	Water / PEG400 / Tween80	Wat PEG Twe
Propylene / Glycol / Tween80	PG Twe	Water / PEG400 / Glycerin	Wat PEG Gly
Propylene Glycol / Glycerin	PG Gly	Water / Propylene Glycol / Tween80	Wat PG Twe
Tween80 / Glycerin	Twe Gly	Water / Propylene Glycol / Glycerin	Wat PG Gly
		Water / Tween80 / Glycerin	Wat Twe Gly
		PEG400 / Propylene Glycol / Tween80	PEG PG Twe
		PEG400 / Propylene Glycol / Glycerin	PEG PG Gly
		PEG400 / Tween80 / Glycerin	PEG Twe Gly
		Propylene Glycol / Tween80 / Glycerin	PG Twe Gly

Figure 2: Solvent combinations (SRI's Level 1 screen) used for the paclitaxel solubility study. (Concentrations: ethanol X%v/v in water; water 100%; PEG400 100%; propylene glycol Y%v/v in water; Tween80 Z%v/v in water, and glycerin A%v/v in water).

ketoconazole, mefenamic acid, and nifedipine are examples of Class II drugs. *In vitro-in vivo* correlation (IVIVC) is usually expected for Class I and Class II drugs.

For Class III drugs, permeability is the rate-limiting step for drug absorption. These drugs exhibit a high variation in the rate and extent of drug absorption. Cimetidine, acyclovir, neomycin B and captopril are examples of Class III drugs.

Class IV drugs exhibit a many characteristics that are problematic for effective oral administration. A decade back, extreme examples of Class IV compounds were an exception rather than the rule, yet today about 10% of the drug candidates under development fall into this category. A well-known example of a Class IV drug is paclitaxel.

PROPRIETARY HIGH-THROUGHPUT SOLUBILITY SCREENING SYSTEM

In recent years, there has been an increase in the drugs that fall into BCS Classes II, III and IV. Of these, Class II and IV pertain to drugs that are poorly water soluble. Currently, more than one-third of the drugs listed in the US Pharmacopoeia are poorly water soluble. Poor solubility leads to significant hurdles in the oral absorption and bioavailability of the drug candidate by decreasing its dissolution rate and membrane permeation.

SRI has developed its own proprietary solvent screening system, which is both economical and fast. This system, which we call the High-Throughput Solubility Screening (HTSS) system, uses minimal drug quantities, since in its early development stages, availability of the drug is limited.

Our proprietary screening system is based on a High-Throughput Liquid Handling Instrument (see figure 1) which is used to prepare the excipient combinations. This instrument is programmed for aspirating and dispensing the solvents/ excipients. A predetermined volume of the cosolvent combinations are dispensed into each well in a microtiter plate. After solvent addition, the plates are shaken in a controlled environment for an extended period. Levels of solubility are read using the baseline turbidity of the excipi-

ent. A software program is used to control the solvent combinations and the analytical outputs. Different level screens of solvent combinations are available to address different formulation options. A stock solution of the drug is prepared either in absolute alcohol or DMSO and added into plates prefilled with the premixed cosolvent combinations. The plates are again shaken and analysed using the turbidity reader. Solubility is assessed by the change in turbidity before and after addition of the drug in the solvent system.

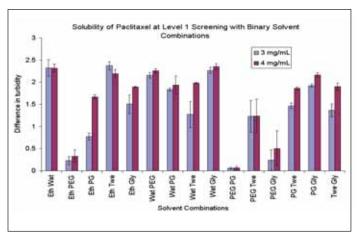


Figure 3: Solubility of paclitaxel (3 and 4mg/mL) with binary combinations of six solvents

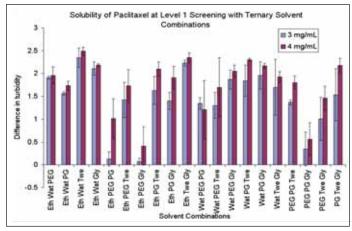


Figure 4: Solubility of paclitaxel (3 and 4 mg/mL) with ternary combinations of six solvents





Figure 5: Top: Ussing absorption unit; Bottom: Slider with the intestinal tissues being mounted between the side-by-side compartmental unit of the Ussing system.

The following is an example of the strategy employed in developing a formulation for a model drug substance, in this case, paclitaxel.

SRI EXAMPLE

A total of 35 solvent combinations (listed in figure 2) were generated by the software using the six solubilising excipients in binary and ternary combinations at a 1:1 and 1:1:1 ratio, respectively. Paclitaxel stock solutions were prepared in absolute ethanol and dispensed into each of the micro-titer plates containing solvent combinations to obtain the desired final concentrations of 3 and 4 mg/mL. Turbidity was measured using the plate reader.

Figures 3 and 4 show the solubility profiles of paclitaxel in both binary and ternary combinations as obtained from the HTSS. In these figures, the longer bars represent more turbidity in the formulation. Based on this analysis, we were able to narrow down a few cosolvent-based formulations options for paclitaxel, which were then tested in a pharmacokinetic study.

PROPRIETARY IN VITRO PERMEATION SYSTEM

Although Class III and Class IV drugs are poorly absorbed substances, in recent times there has been an increase in drug products falling into these categories owing to newer formulation options. Class III compounds are drug substances that are sometimes highly water

soluble, but they do not have the appropriate hydrophobic-hydrophilic balance to be absorbed by the intestinal epithelium.

Developing an oral formulation for these compounds sometimes requires imparting the right hydrophobic-hydrophilic balance to improve the GI-permeation of the drug. In this step in the drug development process, SRI uses an *in vitro* screen known as the Ussing Permeation System to rank-order formulations on the basis of their permeability.

The system at SRI is a modified Ussing system (shown at the top of figure 5) that has multiple sets of side-by-side chambers and is used for *in vitro* studies. Each set consists of two side-by-side diffusion chambers, a heating block for temperature control, needle valves for gas flow adjustment and gas mixing, and Ag/AgCl voltage and current electrodes for measuring transepithelial voltage and for passing current.

Harvested segments of small intestine or colon are mounted on sliders placed between the two horizontal chambers of the modified Ussing system (bottom of figure 5). One of the chambers is exposed to the mucosal side of the intestinal segment and the other to the serosal side.

For drug transport across epithelial membranes of harvested rat small intestine and colon segments in the mucosal-to-serosal (M-to-S) direction, aliquots of buffer are added to both the mucosal and serosal chambers. The buffer in the mucosal chamber is replaced with a solution of the drug formulation. Aliquots of buffer solutions are removed periodically from the serosal chambers and are replaced with equal volumes of fresh warm buffer previously saturated with 100% O₂.

Changes in transepithelial short-circuit current (in micro-Amps) and membrane resistance (in Ohms) as a function of time are monitored continuously during *in vitro* studies to serve as indicators of tissue viability and drug permeability, respectively.

The buffer samples from the receptor chambers are analysed for drug content on a high pressure liquid chromatograph (HPLC). Owing to the expected variation for *in vitro* test conditions and harvested rat intestinal segments, four replicates are used per experimental condition.

Apparent Permeability Coefficient (P_{app}) Calculation:

To compare data obtained from different *in vitro* experiments, the apparent permeability coefficients are calculated using the equation:

$$P_{app} = \frac{dQ / dt}{C_0 \times A}$$

where dQ/dt is the linear appearance rate of mass in the receiver compartment, C_0 is the ini-

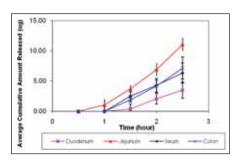


Figure 6: Drug transport in the mucosal-to-serosal direction, across different segments of rat intestine and colon. from buffer.

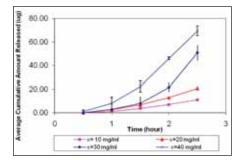


Figure 7: Effect of drug concentration on transport in the mucosal-to-serosal direction across rat jejunum, from buffer.

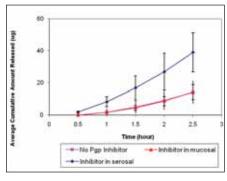


Figure 8: Effect of a known Pgp inhibitor on drug transport in the mucosal-toserosal direction across rat jejunum.

tial solute concentration in the donor compartment, and A is the surface area.²

The following is an example of the strategy employed in developing a formulation for a model BCS Class III drug substance.

SRI EXAMPLE

Figures 6, 7 and 8 illustrate this strategy for a poorly permeable drug substance. The plot between the amount of drug transported through the different regions of the intestine from the mucosal to the serosal side for the drug at a given concentration and formulation is presented in figure 6.

The cumulative amount of drug transported through jejunum fragments of the intestine from the mucosal to the serosal side at increasing drug dose concentrations (in the same formulation), is shown in figure 7.

Figure 8 presents data obtained from using a Pgp (permeability glycoprotein) inhibitor, which was placed either on the mucosal side or on the serosal side, compared with that without at inhibitor. In this figure, the dose, formulation, and kind of intestinal segment used are constant. Similar data compilation continues until all conditions and formulation options are covered.

Based on calculated P_{app} values (data not shown) obtained from in vitro data, the rate of M-to-S transport of drug was found to be highest in jejunum, followed by colon and ileum, and lowest in duodenum (figure 6). Increasing the concentration of drug in the mucosal (donor) chamber increased M-to-S transport of drug in a dose-dependent manner (figure 7). Increased M-to-S transport of drug was observed in vitro, when a known Pgp inhibitor was added to the mucosal or serosal chambers. Addition of a known Pgp inhibitor to the serosal chamber caused a dramatic (three-fold) increase in M-to-S transport of drug. Addition of inhibitor to the mucosal chamber also increased M-to-S transport of drug, but to a lesser extent (figure 8).

The above observations are indicative of the significant role of presystemic elimination processes in poor permeation of drug across the GI tract. The secretory transporters involved may include Pgp, the family of multi-drug-resistance-associated proteins (MRP), and possibly other transporters operating across the GI tract.

SRI's strategy is to study the permeation properties of the drug substance *in vitro* with and without added permeation and solubility enhancers. Experiments are also conducted in the presence and absence of inhibitors of efflux proteins (glycoproteins that pump out the absorbed drugs through independent pathways, resulting in a net decrease in the amount of drug in the serosal side). If the drug substance is found to be influenced by the efflux proteins, the formulation strategy will include inhibition of those proteins.

IN SITU PHARMACOKINETIC DATA BY INTRA-DUODENAL ADMINISTRATION IN RATS

Once the *in vitro* permeation data has been obtained, we determine the pharmacokinetic parameters of the drug in the presence of various formulation additives/permeation enhancers upon intra-duodenal administration

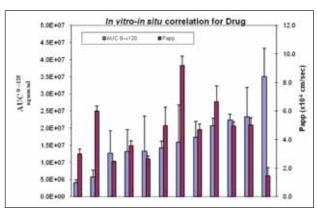


Figure 9: AUC versus Papp values obtained for different drug formulations

in rats. The *in situ* absorption of the drug in the presence of the various additives and enhancers and upon intra-intestinal administration is evaluated in healthy rats. The rats are fasted overnight and anaesthetised so that the abdomen can be opened for isolation of GI tract. Ligatures are placed above the mid-duodenum and below the mid-jejunum, and an intestinal dosing receptacle is created; the drug is then administered into the lumen of intestine by injecting formulation compositions at midduodenum level. Blood samples are collected pre- and post-dose, processed for collection of plasma, and analysed by LC-MS.

Selection of formulation additive combinations is based on the P_{app} values obtained from comparative *in vitro* studies. The absorption of the drug is determined by calculating the area under the plasma drug concentration versus time curve (AUC). Based on the results from *in vitro* and *in situ* studies, AUC values obtained are compared with the corresponding P_{app} values calculated using *in vitro* experimental data. Figure 9 is a bar graph comparing the AUC values with their corresponding P_{app} values for the different compositions studied.

In vitro studies using the Ussing system may be useful for determining the preferential site of drug absorption for BCS Class III/ IV drugs, and for screening and selection of formulation additives as permeation enhancers. In situ studies would further facilitate optimising the concentrations and ratios of additives used, therefore reducing the cost and number of in vivo evaluations for formulation optimisation. In vitro-in situ correlations performed in our laboratories have indicated that it is difficult to predict the in vivo performance of different formulation additives solely based on in vitro transport studies using isolated intestinal segments at higher concentrations of permeation enhancers or when paracellular transport is the mode of drug absorption.

CONCLUSION

Now is the time for integrating highthroughput experimental techniques into the preformulation and formulation steps in the drug discovery process. Companies are screening larger numbers of drug candidates than ever before, and the decision to choose one drug candidate over another has become more complicated because increasing numbers of these drugs fall into BCS Classes II to IV.

The decision to choose a poorly soluble drug over a more soluble counterpart may be required for improved efficacy

and safety, if the formulation can address the solubility issues. In drug discovery, time is of essence, and a fast *in vitro* method is required to screen the drug in the formulation alongside the drug itself.

No longer is it the drug alone, but the drug in combination with its formulation additives, that determine the probability of success for that molecule. SRI International has successfully used the above strategy for many client companies, helping these companies screen out many potential drug candidates at a faster rate. We anticipate that the next few years will see many such timeand resource-saving revolutions in the way we attempt to develop these formulations.

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